

A complex biogeographic history of diversification in Neotropical lancehead pitvipers (Serpentes, Viperidae)

Breno Hamdan^{1,2}  | Thaís B. Guedes^{3,4}  | Paola A. Carrasco^{5,6} | Jane Melville⁷ 

¹Departamento de Genética, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Brazil

²Laboratório de Coleções Biológicas e Biodiversidade, Instituto Vital Brazil, Niterói, Brazil

³Programa de Pós-Graduação em Biodiversidade, Ambiente e Saúde, Centro de Estudos Superiores de Caxias, Universidade Estadual do Maranhão, Caxias, Brazil

⁴Department of Biological and Environmental Sciences and Gothenburg Global Biodiversity Center, University of Gothenburg, Göteborg, Sweden

⁵Facultad de Ciencias Exactas, Físicas y Naturales, Centro de Zoología Aplicada, Universidad Nacional de Córdoba, Córdoba, Argentina

⁶Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), Instituto de Diversidad y Ecología Animal (IDEA), Córdoba, Argentina

⁷Sciences Department, Museum Victoria, Melbourne, Vic., Australia

Correspondence

Thaís B. Guedes, Centro de Estudos Superiores de Caxias, Programa de Pós-Graduação em Biodiversidade, Ambiente e Saúde, Universidade Estadual do Maranhão, Praça Duque de Caxias s/n, 65604-380 Caxias, MA, Brazil.
Email: thaisbguedes@yahoo.com.br

Funding information

Coordenação de Aperfeiçoamento de Pessoal de Nível Superior—CAPES; Fundação de Amparo à Pesquisa do Estado de São Paulo—FAPESP; Universidade Estadual do Maranhão—UEMA; Instituto Vital Brazil—IVB.

Abstract

Based on the literature, we had predicted that the diversification within the Neotropical snake genus *Bothrops* occurred along a latitudinal gradient from north to south, with diversification into unoccupied niches through ecological opportunity, not correlated with geoclimatic events. Using a dated phylogeny and estimating likelihoods of ancestral states at cladogenesis events, we reconstructed ancestral areas and assessed major events of the diversification of *Bothrops* clades, and we also discuss systematic implications for this group. Based on the phylogeny we produced, *B. lojanus* was not considered as part of the genus *Bothrops* since the results recovered this species nested within the *Bothrocophias* clade. We infer that the diversification of the Miocene *Bothrops pictus* and *Bothrops alternatus* clades may be related to the uplift of the western slopes of the Andes and the Argentinian Patagonian Andes, respectively. The Pliocene *Bothrops taeniatus* and *Bothrops osbornei* clades may be related to the uplift of the eastern and northern Andes, respectively. The Plio-Pleistocene *Bothrops neuwiedi* clade may be related to the habitat transitions from a warmer and forested environment to a cooler and open landscape; the *Bothrops jararaca* (i.e. island endemic species) and *Bothrops lanceolatus* clades to over-water dispersal with island speciation; and *Bothrops atrox* clade to the appearance of the Panamanian land bridge. We found that a multitemporal and multidirectional history of diversification may be correlated with geoclimatic and dispersalist events. We argue that the vacant niche hypothesis by itself does not explain *Bothrops* diversification.

KEYWORDS

distribution, diversification, ecological opportunities, molecular phylogeny, Neotropical region, patterns of speciation

1 | INTRODUCTION

The Neotropical region, which corresponds to most of South America, Central America, southern and central Mexico, and the Antilles (Morrone, 2014), comprises the largest biodiversity on Earth (Antonelli & Sanmartín, 2011; Myers, Mittermeier, Mittermeier, Fonseca, & Kent, 2000). Some of the hypotheses on diversification in the Neotropics assert it was due to allopatric speciation and ecological displacement due to the uplift of mountain ranges (Antonelli, Nylander, Persson, & Sanmartín, 2009), to rivers that have acted as prezygotic barrier (Wallace, 1852), to habitats fragmented through Pleistocenic climatic changes (Haffer, 1969), or to a continuous diversification trend, without significant differences between Paleogene, Neogene and Quaternary speciation rates (Bacon, Molnar, Antonelli, Crawford, Montes & Vallejo-Pareja, 2016; Hoorn et al., 2010; Rull, 2011). Finally, some hypotheses argue that intrinsic ecophysiological barriers such as temperature for ectothermic vertebrates (e.g. Hamdan, Pereira, Loss-Oliveira, Rödder, & Schrago, 2017), and rapid diversification due to radiation into niches that were largely unoccupied (Wüster, Salomão, Quijada-Mascareñas, Thorpe, & BBBSP, 2002) contributed to this diversification. The last one implies that when colonizing a habitat with vacant niches, lineages may respond to this “ecological opportunity” by diversifying into several daughter species, each occupying different parts of the ecological space (Schluter, 2000).

Despite numerous attempts to determine the role of historical and ecological factors in the diversification, evolutionary studies of Neotropical groups are still needed to provide a comprehensive picture of the origin of the biodiversity (Lohmann, Bell, Calió, & Winkworth, 2013). Many of these studies focus on groups that are geographically or ecologically restricted (Guarnizo et al., 2016; Smith et al., 2014; Werneck, Leite, Geurgas, & Rodrigues, 2015). Study systems spanning the Neotropics across numerous ecological settings provide a greater ability to test these broad models of diversification.

Venomous snakes of the family Viperidae are a well-known component of the faunal community of the Neotropics, with several species co-occurring in most areas. *Bothrops* (sensu Carrasco, Mattoni, Leynaud, & Scrocchi, 2012), commonly referred to as “lanceheads,” is one of the most speciose genera within the subfamily Crotalinae, with 46 known species widespread in the Neotropical region from Mexico to Argentina (Carrasco et al., 2019; Guedes et al., 2018; Timms et al., 2019; Uetz, Freed, & Hošek, 2018; Wallach, Williams, & Boundy, 2014). Based on phylogenetic data (Fenwick, Gutberlet, Evans, & Parkinson, 2009; Wüster, Salomão, et al., 2002), *Bothrops* can be grouped into six evolutionary clades: *B. alternatus* clade, *B. atrox* clade, *B. jararaca* clade, *B. jararacussu* clade, *B. pictus* clade, *B. newiiedi* clade, and

B. taeniatus clade. First estimates for divergence time of *Bothrops* were around 13 million years ago (Ma) (Hedges, 1996; Savage, 1966). The study of Wüster, Salomão, et al. (2002), who were the first to amass a significant volume of mitochondrial DNA data of the genus, suggested that the ancestor of *Bothrops* might have colonized South America during the Miocene, c. 23–10 Ma. Among latest studies, some have estimated similar divergence time for bothropoids (*Bothrops* + *Bothrocophias*) (Alencar et al., 2016), while others estimated a more recent period of diversification for the group (around 13 Ma or less) (Fenwick, Greene & Parkinson, 2012; Wüster, Peppin, Pook, & Walker, 2008).

Bothrops distribution patterns, high richness and the wide variety of niches they occupy provide an ideal study system for assessing general diversification patterns, testing and refining biogeographic hypotheses in the Neotropics. At first, a biogeographic study suggested that the ancestor of modern *Bothrops* dispersed into South America from the north (Parkinson, Campbell, & Chippindale, 2002). Then, Wüster, Salomão, et al. (2002) reported that the adaptive radiation of *Bothrops* into niches devoid of other viperids might have led to their rapid and great diversification through ecological opportunities. However, this study used 26 of 43 described species at that time and did not perform biogeographic analysis.

We used a data set of mitochondrial DNA, incorporating four gene regions (total 2,058 bp), with sequences from 34 species of *Bothrops* to produce a calibrated phylogeny, and reconstruct the diversification of *Bothrops* clades across the Neotropics. Based on the vacant niches hypothesis, we predict that this geographic pattern of speciation was mostly unidirectional, which means that *Bothrops* had a chronologically southward movement and that the diversification history was not correlated with geoclimatic events. We then use this new information to assess the implications for the systematics of this group.

2 | MATERIAL AND METHODS

2.1 | Study system: genus *Bothrops* (Viperidae: Crotalinae)

The *Bothrops* genus is distributed across the Neotropics. The genus comprises abundant and widely distributed species (e.g. *B. asper* and *B. atrox*) as well as rare and/or restricted species (e.g. *B. pirajai* and *B. muriciensis*). Species occur from sea level to 3,500 m (*B. jonathani*, *B. ammodytoides*) (Campbell & Lamar, 2004; Carrasco, Harvey, & Muñoz Saravia, 2009; Carrasco, Leynaud, & Scrocchi, 2010; Harvey, 1994; Lirada-Silva et al., 2009; Wallach et al., 2014). Most species are continental, terrestrial, and inhabited forested habitats (e.g. *B. atrox*, *B. jararaca* and *B. leucurus*). However, some are arboreal in rainforests (e.g. *B. bilineatus*), and some inhabit

open landscapes (e.g. *B. erythromelas*) (Barbo et al., 2016; Barbo, Grazziotin, Sazima, Martins, & Sawaya, 2012; Campbell & Lamar, 2004; Guedes, Nogueira, & Marques, 2014; Guedes et al., 2018; Turci, Albuquerque, Bernarde, & Miranda, 2009).

Some evolutionary relationships remain controversial, including the status of the Andean *B. lojanus*, reported as incertae sedis by Carrasco et al. (2012) and found to be nested within the genus *Bothrocophias* by Arteaga et al. (2016) and Alencar et al. (2016). Fenwick et al. (2009) proposed a rearrangement in the classification of *Bothrops* recognizing the genera *Bothriopsis*, *Bothropoides* and *Rhinocerothis*, subsequently considered synonyms of *Bothrops* by Carrasco et al. (2012). This synonymy was supported by Alencar et al. (2016) and has also been adopted in the current study.

2.2 | Taxon sampling and data acquisition

We sampled 34 species of *Bothrops* representative of the major clades within the genus, as well as 51 species from 23 other genera than *Bothrops* and the lizard *Iguana iguana* for rooting and allowing a greater number of calibration points. DNA sequences, incorporating four mtDNA genes (12S, 16S, ND4, CYTB), were downloaded from GenBank for each species, except for *B. leucurus*, which was sequenced as part of the current study. Alcohol-preserved tissue samples of *B. leucurus* were extracted from liver using the Qiagen DNeasy kit. Sequence amplification via polymerase chain reaction (PCR) was performed using gene-specific primers as follows: 12S, L12 “5-CGCCAAAYA ACTACGAG-3” (Vidal, Lecointre, Vié, & Gasc, 1997) and H1557 “5-GTACACTTACCTTGTTACGACTT-3” (Knight & Mindell, 1994); 16S, 16Sar “5-CGCCTGTTTATCAAAA CAT-3” and 16Sbr “5-CTCCGGTCTGAACTCAGATCAG TAGG-3” (Palumbi et al., 1991); ND4 ND4f, “5-CACCTA TGACTACCAAAGCTCATGTAGAAGC-3” and ND4fr, “5-TTCTATCACTTGGATTTGCACCA-3” (Arévalo, Davis, & Sites, 1994); and CYTB LI4724, “5-TGACTTGAARAAC CAYCGTTG-3” and H15915, “5-TGAGAAGTTTTCYGGG TCRTT-3” (Irwin, Kocher, & Wilson, 1991; Kocher et al., 1989). The GenBank accession number for sequences are provided in Appendix S1.

Ribosomal 12S and 16S markers were aligned with MAFFT software (Kato & Standley, 2013) using the Q-INS-i algorithm, which considers the secondary structure of RNA. Sequences of CYTB and ND4 were aligned using the default settings of MAFFT. Minor adjustments were done manually. The final data set consisted of 2,058 bp from the following four genes: two mitochondrial protein-coding genes, cytochrome b (CYTB, 635 bp; 86 taxa) and NADH subunit 4 (ND4, 624 bp; 86 taxa); and two non-coding ribosomal genes, 12S (333 bp; 74 taxa) and 16S (478 bp; 76 taxa).

2.3 | Supermatrix tree analyses and time of diversification

The GTR+G+I model of sequence evolution was chosen for each partition independently by jModelTest using both AIC and BIC criteria (Darriba, Taboada, Doallo, & Posada, 2012; Guindon & Gascuel, 2003). We inferred maximum likelihood topologies using RAxML 8.2.0 on the CIPRES science gateway (Miller, Pfeiffer, & Schwartz, 2010) and assessed the statistical confidence of branches by 1,000 bootstrap replicates (Felsenstein, 1985). We performed Bayesian Inference in MrBayes 3.1.2 software and ran the Markov chain Monte Carlo (MCMC) algorithm for 80,000,000 generations using three independent runs with four chains each. Chains were sampled every 1,000th generation. We applied a burn-in of 25% and evaluated effective sample size values using TRACER v.1.5.

A time tree was inferred using the RelTime method in MEGA 7 (Tamura et al., 2012) and the GTR model. The timetree was computed using eight calibration constraints. A discrete Gamma distribution was used to model evolutionary rate differences among sites (four categories (+G, parameter = 0.6745)). The rate variation model allowed for some sites to be evolutionarily invariable ([+I], 33.21% sites).

The calibration points used to convert relative times into absolute time estimates of divergence times were based on snake fossils (Buffetaut, Marandat, & Sigé, 1986; George & Vincent, 1977; Head, 2015; Head, Mahlow, & Müller, 2016; Parmley & Holman, 2007; Parmley & Hunter, 2010; White, 1942) and were defined as follow: (a) the split between “*Eryx–Boinae*” between 58.0 and 64.0 (hard minimum–hard maximum ages) million years ago (Ma), which corresponds to the divergence between “*Erycinae–Boinae*”; (b) the split between “*Boa–(Epicrates + Eunectes)*” was constrained to be older than 20.43, which corresponds to the minimum age of the oldest *Boa* fossil (stem group age of *Boinae*); (c) the split between “*Eunectes–Epicrates*” was constrained to be older than 12.3 Ma, which corresponds to the minimum age of the oldest *Eunectes* fossil; (d) the split between “*Oxyuranus–Laticauda*” was constrained to be older than 10.0 Ma, which corresponds to the minimum age of the oldest *Oxyuraninae* fossil (minimum divergence between *Laticauda* and *Oxyuraninae*); (e) the split between “*Elapoidea–Colubridae*” was constrained to be older than 30.8 Ma, which corresponds to the minimum age of divergence of crown colubroid lineages; (f) the split between “*Booidea–Caenophidia*” was constrained to be older than 72.1, which corresponds to the oldest fossil assigned to *Caenophidia* crown group; (g) the split between “*Viperidae–(Elapoidea + Colubridae)*” was constrained based on the oldest fossil of this clade, an *Elapidae* of 48.6 Ma; and (viii) the split between “*Crotalus–Sistrurus*” was constrained to be older than 10.3 Ma, which corresponds to the oldest *Sistrurus* fossil.

There has been much critical attention towards dated phylogeny, especially on the use of calibration, which is the

rate-determining step in every molecular clock analysis. To minimize the issue on distinguishing good from bad calibrations, we run a complementary timetree analysis using Viperidae species and only *Sistrurus* fossil as the closest constraint. The results are placed in Appendix S5.

2.4 | Historical biogeographic analysis

We sought to identify factors or processes related to shaping the composition and distribution of *Bothrops* major groups over time. Thus, the posterior distribution of divergence times of the *Bothrops* clades was then compared with past geoclimatic events reported in the specific literature.

Another goal was to infer the ancestral areas of clades by using the retrieved timetree and test whether *Bothrops* had a chronologically southward movement and that the diversification history was not correlated with geoclimatic events. We inferred the historical biogeography of *Bothrops* based on RAxML topology due to the high resolution and bootstrap values and, when appropriate, the age nodes from Mega 7, RelTime method. We assigned the distribution of the species to the following pre-defined Neotropical areas (according to Morrone, 2014): Antillean subregion: Antilles or West Indies (Greater and Lesser Antilles) and the Bahamas Islands, including the Bahamas, Cuban, Cayman Islands, Jamaica, Hispaniola, Puerto Rico and Lesser Antilles provinces; Brazilian subregion: Southern and central Mexico, Central America and north-western South America including the Mesoamerican, Pacific, Boreal Brazilian and Southwestern Amazonian dominions; Chacoan subregion: south-eastern South America including the Southeastern Amazonian, Chacoan and Parana dominions; and South American Transition Zone: highlands of the Andes between western Venezuela and northern Chile and central western Argentina, including the Paramo, Desert, Puna, Atacama, Prepuna and Monte provinces. The geographic distribution of each species of *Bothrops* was obtained from scientific literature (Campbell & Lamar, 1989, 2004; Carrasco et al., 2012; Fenker, Tedeschi, Pyron, & Nogueira, 2014; Silva & Rodrigues, 2008; Uetz et al., 2018). Detailed information on the assignment of each species of *Bothrops* in the Neotropical areas adopted is provided in Appendix S2.

We reconstructed the ancestral areas for each node in the phylogeny of *Bothrops* using the Reconstruct Ancestral States in Phylogenies software (RASP; Yu, Harris, Blair, & He, 2015). We chose the Dispersal–Extinction–Cladogenesis model for geographic range evolution (DEC; Ree, Moore, Webb, & Donoghue, 2005), for biogeographic analysis. DEC specifies instantaneous transition rates between ranges along phylogenetic branches and applies these rates to estimate likelihoods of ancestral states (range inheritance scenarios) at cladogenesis events. We limited the maximum number of

regions in ancestral areas to two. Based on results from our phylogeny, *Bothrops lojanus* was not considered as part of *Bothrops* in our biogeographical analyses given that the results recovered the species nested within the *Bothrocophias* clade. This grouping is consistent with the results reported by Arteaga et al. (2016) and Alencar et al. (2016).

3 | RESULTS

3.1 | Phylogenetic relationships

The concatenated tree topologies inferred using ML and BI were highly congruent and generally well resolved. The tree estimated in RaxML showed the best topological resolution. Clades recovered in both ML and Bayesian (Appendix S3) analyses are indicated in Figure 1 with their respective statistical support.

Bothrops lojanus is nested within the genus *Bothrocophias*, recovering *Bothrops* as paraphyletic. *Bothrops pictus* was recovered as the sister group of all remaining representatives of the genus (Figure 1 and Appendix S3). Except for *B. lojanus*, all species of *Bothrops* formed a well-supported monophyletic clade. Thus, based on these results, we herein define the following *Bothrops* clades to be used in the diversification and biogeographic analyses: (a) *B. pictus* clade (contains only *B. pictus*); (b) *B. taeniatus* clade (*B. taeniatus*, *B. pulchra*, *B. bilineatus* and *B. chloromelas*); (c) *B. jararacussu* clade (*B. jararacussu* and *B. brazili*); (d) *B. osbornei* clade (*B. osbornei* and *B. punctatus*); (e) *B. lanceolatus* clade (*B. lanceolatus* and *B. caribbaeus*); (f) *B. atrox* clade (*B. atrox*, *B. asper*, *B. moojeni*, *B. colombiensis*, *B. isabellae*, *B. marajoensis* and *B. leucurus*); (g) *B. jararaca* clade (*B. jararaca* and *B. insularis*); (h) *B. neuwiedi* clade (*B. neuwiedi*, *B. erythromelas*, *B. lutzi*, *B. mattogrossensis*, *B. pubescens*, *B. diporus*, *B. marmoratus* and *B. pauloensis*); and (i) *B. alternatus* clade (*B. alternatus*, *B. ammodytoides*, *B. fonsecai*, *B. cotiara* and *B. itapetingae*).

3.2 | Diversification and patterns of speciation of *Bothrops*

Overall, most *Bothrops* clades are widely distributed in the Neotropical areas adopted here. Fourteen species are restricted to the Chacoan subregion (highest species richness), eight in the Brazilian subregion, two in the Antilles subregion and five to the South American Transition Zone. Five species were assigned to occur in both Chacoan + Brazilian subregions, one in the Chacoan subregion + South American Transition Zone and one in the Brazilian subregion + South American Transition Zone. The *B. jararaca* clade is restricted to the Chacoan subregion, while *B. lanceolatus* clade is exclusive to the Antilles subregion and *B. pictus* to the South American Transition Zone (see Appendix S2).

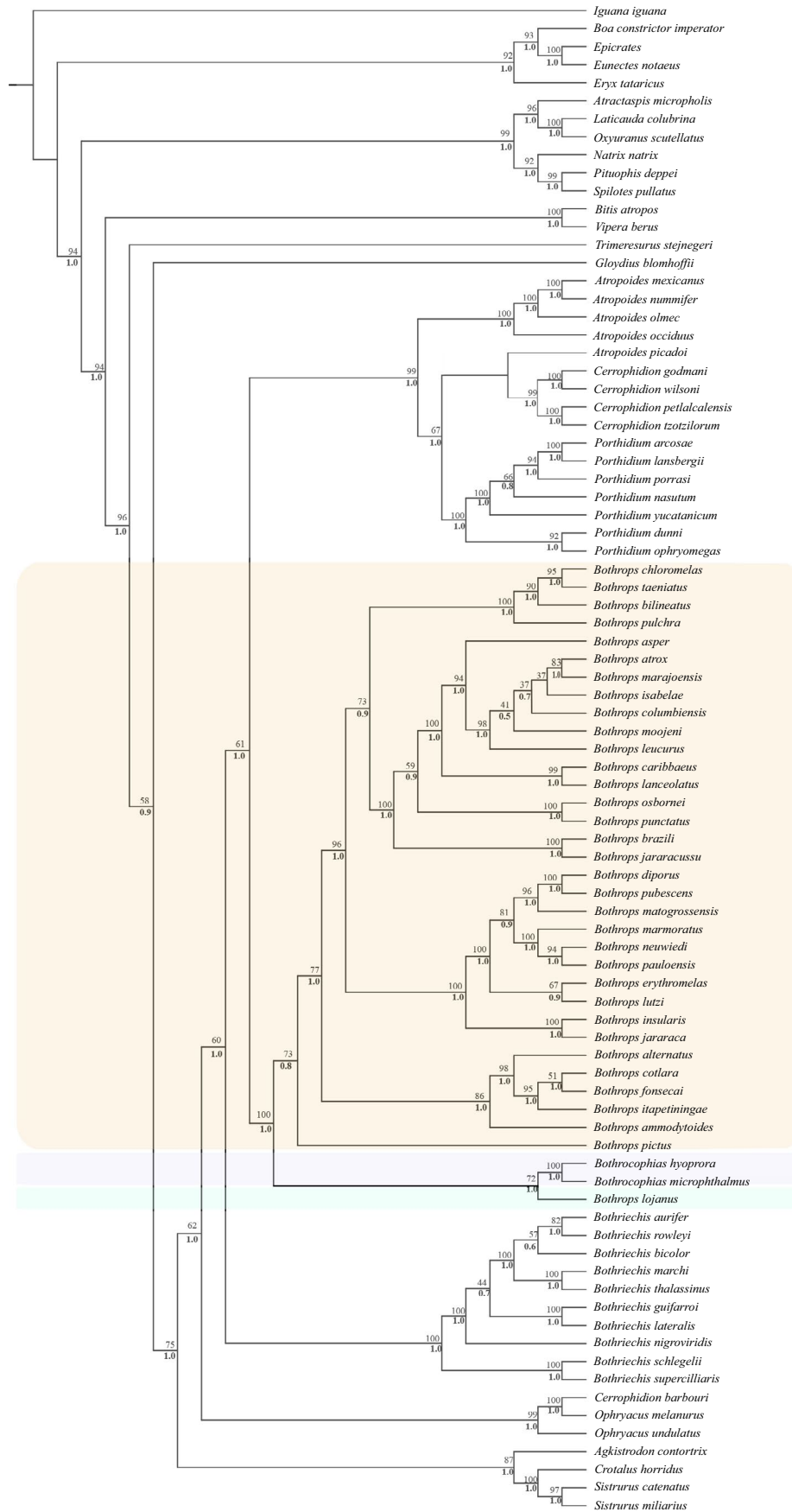


FIGURE 1 Phylogenetic relationships of the species of the genus *Bothrops* based on maximum likelihood inferred in RaxML 8.2.0. Numbers on branches indicate the bootstrap support (not bold) and posterior probability (bold)

Our results show that the most likely ancestral scenario for the diversification of the genus *Bothrops* (nodes 68–71; Figure 2) was within mainland of the South American Transition Zone + Chacoan subregion during the Miocene *c.* 12.03–11.08 Ma (95% divtime: 0.14–32.03). The *B. pictus* clade is supported as having diversified in the South American Transition Zone, splitting from the common ancestor of all other *Bothrops* species (node 67) during the Miocene *c.* 11.08 Ma (95% divtime: 1.91–30.95 Ma).

Results suggest that the *B. alternatus* clade (node 66) has evolved within the Chacoan subregion during the Miocene *c.* 8.81 Ma (95% divtime: 0.29–26.17 Ma). *Bothrops amodytoides*, the sister group to the other representatives of this clade, is the only taxon distributed in both Chacoan and South American Transition Zone areas.

The *B. neuwiedi* clade (node 60) is inferred to have diversified from a common ancestor (node 61) shared with the *B. jararaca* clade, both restricted to the Chacoan subregion. The diversification of the *B. neuwiedi* clade was found to take place during the Pliocene *c.* 3.04 Ma (95% divtime: 0.2–9.25 Ma), while the diversification of the *B. jararaca* clade (node 53) was reconstructed to the Chacoan subregion from an ancestor (node 61) restricted to this region during the Plio/Pleistocene transition *c.* 2.37 Ma (95% divtime: 0.3–7.66 Ma).

The best-supported scenario for the diversification of the *B. taeniatus* clade (node 51) involves the Brazilian subregion + South American Transition Zone during the Pliocene *c.* 5.47 Ma (95% divtime: 0.10–16.77 Ma). The diversification of the *B. jararacussu* clade (node 47) occurred at the Chacoan + Brazilian subregions in the Pleistocene *c.* 2.68 Ma (95% divtime: 0.2–8.66 Ma). The diversification of the *B. osbornei* clade (node 45) involved the Brazilian + South American Transition Zone areas during the Pliocene and posterior speciation from the ancestor node 46 *c.* 3.64 Ma (95% divtime: 0.1–11.9 Ma). The *B. lanceolatus* clade (node 43) is inferred to have diversified in the Antillean subregion from an ancestor that inhabited the Brazilian subregion during the Plio/Pleistocene transition *c.* 1.91 Ma (95% divtime: 0.2–6.43 Ma).

Finally, our results support the diversification of the *B. atrox* clade (node 42) in the Chacoan + Brazilian subregions during the Plio-Pleistocene *c.* 3.02–2.32 Ma (95% divtime: 0.5–7.59 Ma).

4 | DISCUSSION

4.1 | Early diversification—Pliocene/Miocene orogenic events

The crown nodes in *Bothrops* range from the middle Pleistocene *c.* 0.54 Ma, Quaternary (split *B. diporus* + *B. pubescens*) to 11.08 Ma between Middle and Late Miocene, Neogene (*Bothrops* root). A Miocene occurrence of viperids

at South America is supported by the oldest fossil record of the family in the southern continent, a viperid vertebra reported for the late Miocene Cerro Azul Formation at Caleufú, in La Pampa Province, central Argentina (Albino & Montalvo, 2006). The oldest fossil record for *Bothrops* corresponds to a compound bone exhumed from sediments attributed to the Ensenada Formation (La Plata, Buenos Aires Province, Argentina) from the Early to Middle Pleistocene (0.915–1.01 Ma), which indicate that the cladogenetic events that led to the extant *Bothrops* occurred before 1 Ma (Scanferla & Nenda, 2005).

The uplift of the Andes, which began in the late Oligocene to early Miocene (*c.* 23 Ma) with the formation of the highest peaks during the late Middle Miocene (*c.* 11–12 Ma) (Folguera et al., 2011; Hoorn et al., 2010), coincides with the age estimates for the diversification of *Bothrops*. This continuous strip of mountain ranges that separates the rest of the continent from the narrow Pacific coastal region potentially acted as an effective moisture barrier (Aragon et al., 2011), leading to vicariance speciation, while also presenting new opportunities for ecological adaptation and dispersal (Antonelli et al., 2009; Elias et al., 2009). In the late middle Miocene (*c.* 12.9–11.8 Ma), the eastern Cordillera started developing, extending for almost 9,000 km along the western coast of South America. Thus, it is possible that the ancestor of *Bothrops* was widely distributed and was then splitted by the Andean uplift, which probably also played an important role in the diversification events within *Bothrops* during the Miocene. According to Martins, Araujo, Sawaya, and Nunes (2001), it is possible that the ancestor of *Bothrops* was a small snake, with stout body, and had terrestrial habits.

According to our results, *B. pictus* is the sister group to all *Bothrops* species. The species distribution is restricted to Peru, from the western slopes of the Andes to the Pacific coast, from La Libertad to Arequipa Departments, inhabiting arid to semiarid coastal foothill environments, river valleys and dry western Andean slopes (Campbell & Lamar, 2004; P. A. Carrasco & P. J. Venegas, unpublished data). There is evidence that in the late Miocene the Andean uplift continued to play a role in the diversification and speciation within *Bothrops*. The divergence of *B. amodytoides* from all *B. alternatus* clade *c.* 8.81 Ma, supports the hypothesis of its occurrence during Pliocene/Miocene orogenic events, potentially due to the uplift of the Patagonian Andes. The current distribution of *B. amodytoides*, endemic to Argentina, includes areas of the Puna, Prepuna and Monte (part of the South American Transition Zone) and extends to areas in the dry Chaco, Espinal and Pampas, from sea level to altitudes of more than 3,000 m, reaching the highest austral latitudes among snakes. This distribution pattern suggests a south-east expansion from an Andean ancestor, as has been proposed previously (Carrasco et al., 2010; Werman, 2005).

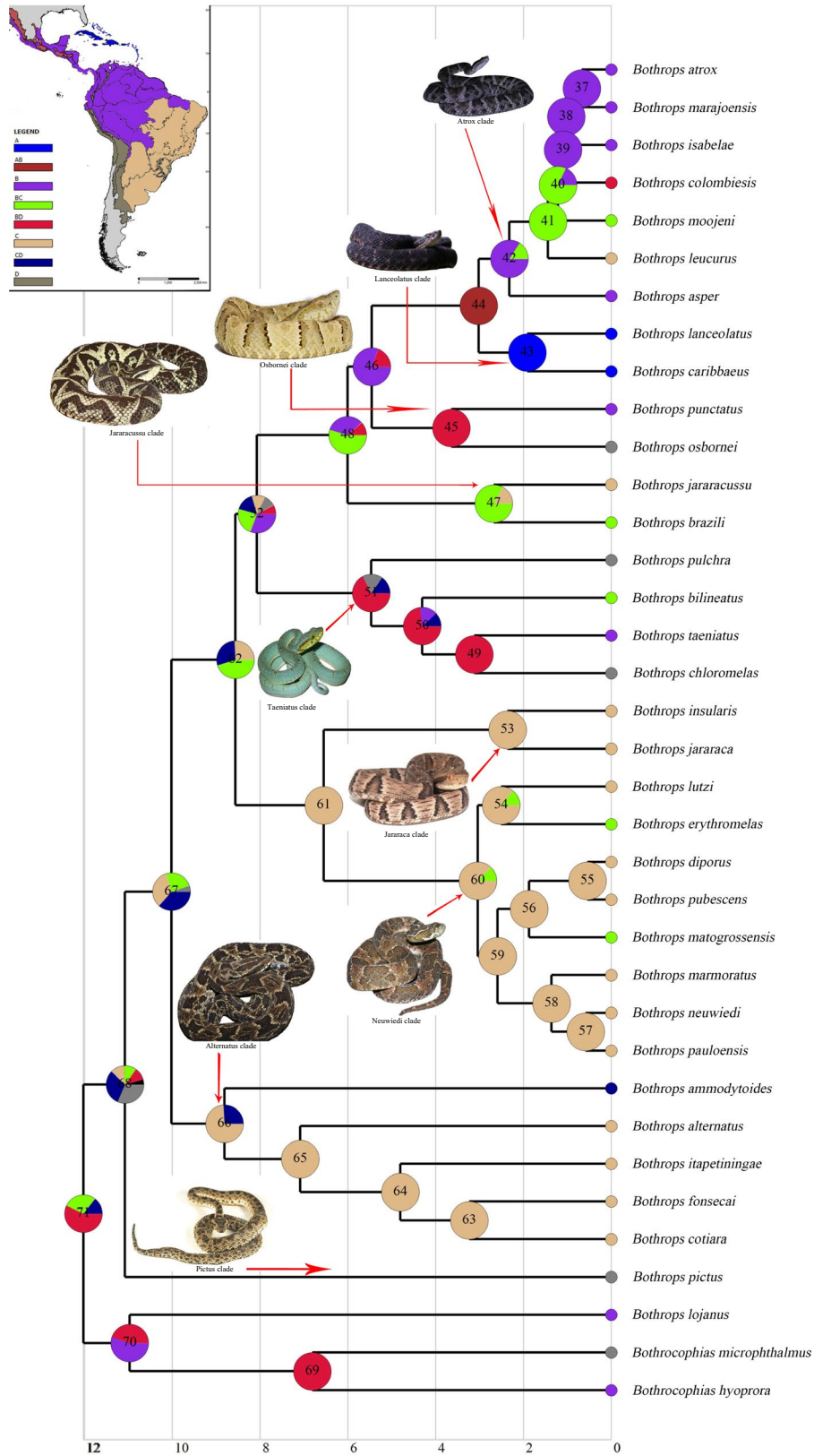


FIGURE 2 Biogeographic history of the snakes genus *Bothrops* through temporal and geographical aspects of diversification from DEC analysis. The bar at the bottom side gives the absolute time scale in millions of years before the present. The modern range for each taxon is colour-coded and is drawn on the terminal lineages before each taxon's name. Pie charts at internal nodes represent the marginal probabilities for each alternative ancestral area. Only ancestral areas for basal nodes are shown. Codes. Chacoan subregion: C; Brazilian subregion: B; South American Transition Zone: D; Antillean subregion: A. Photograph credits: Rodrigo Tinoco (Jararaca clade), Marco Freitas (Taeniatus clade and Jararacussu clade), Omar Torres-Carvajal (Osbornei clade), Germán Chávez (Pictus clade), Catherine Godefroid (Lanceolatus clade)

More recent diversification during the Pliocene led to the formation of the *Bothrops taeniatus*, *B. osbornei* and *B. jararacussu* clades, a period in which Andean elevations rapidly increased by 60% between 2 and 5 Ma, reaching modern elevations by around 2.7 Ma (Gregory-Wodzicki, 2000). This may have led to heterogeneity at local and regional scales, potentially enhancing environmental opportunity for ecological divergence and speciation (Werneck, 2011). The Pliocene rapid uplift of the Eastern Cordillera of the Andes at rates of 0.6–3 mm/year (Gregory-Wodzicki, 2000) may have played a role in the diversification of the *B. taeniatus* clade (node 51), with the speciation of *B. pulchra* c. 5.47 Ma along the South America Transition Zone. *Bothrops pulchra* currently inhabits montane temperate forests in the eastern slopes of the Andes, from southern Colombia to northern Peru (Campbell & Lamar, 2004). The diversification of *B. osbornei* clade (node 45), with the speciation of *B. osbornei* at the South American Transition Zone and *B. punctatus* at the Brazilian subregion, seems to be coupled to the uplifts of the northern Andes c. 5.46 Ma (node 46) (Hoorn et al., 2010). Currently, *B. osbornei* inhabits montane forests in the Pacific slopes of the Andes in Ecuador, and *B. punctatus* inhabits coastal plains and montane forests from eastern Panama to the Pacific coast and Pacific slopes of the Andes in Colombia and Ecuador (Campbell & Lamar, 2004; Cisneros-Heredia, Borja, Proaño, & Touzet, 2006; Ospina-L, 2017). The evolution of the *B. jararacussu* clade is the subject of an ongoing research, and it will not be discussed in details here.

4.2 | Plio-Pleistocene scenario of climatic oscillation, Panamanian land bridge uplift and dispersal over water

The Plio-Pleistocene was a period of transition from humid and warmer forest-like habitats to cooler and drier savannah-like habitats (Hooghiemstra & Cleef, 1995). This period was characterized by pronounced dry seasons (Jacobs, 2004) associated with drastic lowering of global temperatures and increasing aridity, resulting in the replacement of lowland rainforests by savannah woodlands (Plana, 2004). From that time, environmental shifts occurred during the entire Quaternary (2.588 Ma to present) and were linked to climatic changes (temperature and aridity oscillations). It is possible that these dramatic changes in climate and habitats shaped the diversification and speciation patterns of younger clades in *Bothrops*. The influence of Plio-Pleistocene events related to extreme environmental heterogeneity have been reported in a range of Neotropical fauna, including snakes of the genus *Pituophis* (Bryson, García-Vázquez, & Riddle, 2011), mosquitoes (Conn & Mirabello, 2007) and birds (Grau, Pereira, Silveira, Wajntal, & Höfling, 2005).

The role of such climate shifts in the diversification of *Bothrops* is denoted in the common ancestor of *B. neuwiedi* and *B. jararaca* clades (node 61), which seem to have diversified in the late Miocene c. 8.56–6.55 Ma, predominantly in the Chacoan subregion with few subsequent dispersal events towards the Brazilian subregion (nodes 54 and 56) during the Pleistocene (c. 2.6–1.95 Ma). The diversification of the *B. neuwiedi* clade (node 60) took place with the split of two major clades (*B. erythromelas* + *B. lutzii*) and a clade that comprises the rest of the species of the group. Machado, Silva, and Silva (2014) highlighted the importance of Neogene events in the diversification of the *B. neuwiedi* clade and proposed that Quaternary climate fluctuations were responsible for the diversification within the clade. During Pleistocene climatic shifts associated with glaciation, areas of moister vegetation types (such as rainforest) and their populations contracted and became isolated, surrounded by drier vegetation such as savannah. During warmer, wetter periods, these vegetation islands (refugia) and their populations expanded. The contractions and expansions occurred cyclically and are thought to have resulted in diversification (Haffer, 1969).

The *Bothrops jararaca* clade (node 53) diversified within the Chacoan subregion with the split of mainland *B. jararaca* and the insular species *B. insularis* c. 2.37 Ma, values that are much older than the c. 10,000 years previously reported (Grazziotin, Monzel, Echeverrigaray, & Bonatto, 2006), but in agreement with the recent literature (e.g. Alencar et al., 2016). Geologic estimates of maximum Pliocene sea level ranged from +5 to +40 m relative to present, with +25 m typically used by the modelling community (Raymo et al., 2009). Thus, considering that there is no strong evidence of sea-level fluctuation by that time, we infer that the diversification of the *B. jararaca* clade may be due to a dispersal event, from mainland populations to islands through dispersal over water. Another existing island endemic species of the *B. jararaca* clade such as *B. alcatraz*, *B. otavioi* and *B. sazimai* may support this hypothesis. Nevertheless, the shared mitochondrial haplotypes between the continental and island forms (Grazziotin et al., 2006) as well as the other terrestrial species found on the Queimada Grande island may suggest secondary contact by connection via land, perhaps mediated by more recent cycles of sea-level transgressions.

In addition to climate and vegetation transition during the Plio-Pleistocene, the uplift of the Isthmus of Panama linking North and South America during the late Pliocene, led to unprecedented ecological and evolutionary consequences for previously isolated biotas (Simpson, 1980). Among the many evolutionary inferences gleaned from the mammalian fossil record are the following: (a) immigrant taxon appeared on each continent soon after the formation of the Panamanian Isthmus, (b) an early wave of xeric-adapted species was followed by a second wave of mesic-adapted species; and (c)

most exchanges had ceased by the onset of the mid-Pleistocene (Webb, 1976).

The *B. atrox* clade (node 42) started its diversification c. 3.02–2.32 Ma. Our results support the hypothesis that *B. asper* was the first lineage to diversify, which matches with the appearance of the Panamanian land bridge, allowing this taxon to be the only *Bothrops* species to have reached Central America. *Bothrops asper* currently occurs in the forests of Central and northern South America (see Wüster, Salomão, Duckett, Thorpe, & BBBSP, 1999; Wüster, Salomão, et al., 2002). Speciation between North, Central and South America c. 3 Ma has been reported in many faunal clades (Marshall, Webb, Sepkoski, & Raup, 1982), suggesting the formation of the Isthmus of Panama may explain the early diversification of the *B. atrox* clade (see Parkinson et al., 2002). Our estimates of the diversification of the *B. atrox* clade during the Plio-Pleistocene, together with the enormous variety of habitats occupied by its species—from rain forests (e.g. *B. leucurus* and *B. atrox*) to savannas (e.g. *B. moojeni*)—are suggestive of the Refuge Hypothesis (Haffer, 1969, 2008) as an explanation for subclade diversification. Finally, the ancestor of the *B. atrox*–*B. lanceolatus* clade may have dispersed over water to the islands of the Antillean subregion during the Pleistocene and diversified into the species of the *B. lanceolatus* clade, as previously reported by Wüster, Thorpe, et al. (2002). Currently, *B. lanceolatus* and *B. caribbaeus* inhabit the Martinique and Santa Lucía Islands, respectively, in the Lesser Antilles (Wüster, Thorpe, et al., 2002). Saldarriaga-Córdoba, Parkinson, Daza, Wüster, and Sasa (2017) consider an alternative hypothesis to explain the allopatric distribution of *B. atrox* group and *B. asper* lineages on each side of the Eastern Andes Cordillera, suggesting that the final uplift of this mountain range played a significant role in the cladogenesis of these lanceheads. They report that *B. asper* stock would diverge to the west of the Eastern Andes, possible within the foothills along the Pacific coast of northern South America and that the uplift of the Panamanian land bridge would have played a role in the diversification within *asper* lineages.

Overall, our data are in congruence with the history of diversification found for snakes of the genus *Lachesis* (Zamudio & Greene, 1997) and *Corallus* (Colston et al., 2013), frogs (Weigt, Crawford, Rand, & Ryan, 2005), birds (Ribas, Aleixo, Nogueira, Miyaki, & Cracraft, 2012), bees (Ramírez, Roubik, Skov, & Pierce, 2010), butterflies (Brower, 1994) and plants (Antonelli et al., 2009; Dick, Abdul-Salim, & Bermingham, 2003), which would have involved geoclimatic events during the Neogene, dispersals from South America to Central America via the Panamanian land bridge and over-water dispersals to the Antilles during the late Neogene. This congruent biogeographic patterns provide empirically robust diversification hypotheses and support the history of speciation for the clades of *Bothrops* presented herein.

The choice of calibration points is a critical part of diversification studies (Inoue, Donoghue, & Yang, 2010). The complementary timetree analysis using Viperidae species and only *Sistrurus* fossil as the family closest constraint presented younger ages overall, different age values from those found in the literature (e.g. Alencar et al., 2016) and wider confidence intervals (Appendix S5). This is consistent with literature observations on mean substitution rate and coefficient of variation in rates. Sauquet et al. (2012) suggest that, even when suitable internal age constraints for the in-group are available, more accurate age estimates can be obtained by combining both in-group and out-group calibrations. Thus, we have decided to discuss the results for the tree calibrated with all the fossil constrains (Appendix S4, Table 1).

4.3 | Systematic implications

The phylogenetic relationships among the major clades within *Bothrops* shown by our results are in agreement with previous studies (Alencar et al., 2016; Carrasco et al., 2012); these clades are well supported by both molecular and morphological evidence. Our results also support the monophyly of *Bothrops* + *Bothrocophias*; *Bothrocophias* being a genus composed of six species whose distribution is mainly Andean. However, some phylogenetic relationships at the point of divergence between both lineages are not yet sufficiently clear.

Our analyses recovered *Bothrops lojanus* nested within *Bothrocophias*; this relationship was well supported by bootstrap values and is in agreement with previous results (e.g. Alencar et al., 2016). The systematic position of *B. lojanus* was enigmatic until it was included in phylogenetic analyses, showing different results. *Bothrops lojanus* is an Andean species known from a restricted area in southern Ecuador and northern Peru (Campbell & Lamar, 2004; Carrasco, Venegas, Chaparro, & Scrocchi, 2016). Based on morphological evidence, the species has been recovered as the sister taxon of the remaining *Bothrops* (Fenwick et al., 2009) or closely related to *B. pictus* and *B. ammodytoides* (Carrasco et al., 2012). Phylogenies based on mtDNA (Alencar et al., 2016; Arteaga et al., 2016; the present study) recovered the species basal to or rooted within *Bothrocophias*, regarding *Bothrops* paraphyletic. A recent phylogenetic study combining molecular and morphological evidence recovered *B. lojanus* in a position that led *Bothrocophias* to be paraphyletic (Carrasco et al., 2019). Hence, although undoubtedly *B. lojanus* is a basal member of the *Bothrops* + *Bothrocophias* radiation, it may still be considered an “unstable” taxon whose different positions regarding both genera will likely lead to some taxonomic rearrangement. To clarify the phylogenetic position of the Andean *B. lojanus* is crucial for a better understanding of the geographical and historical patterns of divergence of the sister lineages *Bothrops* and *Bothrocophias*, one of the aims of ongoing research projects.

<i>Bothrops</i> clades	Ancestral area	Divergence times mean [95% CI]	Putative events (references)
<i>B. pictus</i>	SATZ	11.08 [1.91–30.95]	Uplift of the Andes (Folguera et al., 2011; Hoorn et al., 2010)
<i>B. alternatus</i>	CS	8.81 [0.29–26.17]	Uplift of the Patagonian Andes (Carrasco et al., 2010; Werman, 2005)
<i>B. taeniatus</i>	BS + SATZ	5.47 [0.10–16.77]	Rapid uplift of the Eastern Cordillera of Andes (Gregory-Wodzicki, 2000)
<i>B. osbornei</i>	BS + SATZ	3.64 [0.1–11.9]	Uplifts of the Northern Andes (Hoorn et al., 2010)
<i>B. neuwiedi</i>	CS	3.04 [0.2–9.25 Ma]	Climate and vegetation changes (e.g. Machado et al., 2014)
<i>B. jararaca</i>	CS	2.37 [0.3–7.66]	Sea-level fluctuations (e.g. Graziotin et al., 2006) and dispersal over water
<i>B. atrox</i>	CS + BS	2.32 [0.5–7.59]	Appearance of the Panamanian land bridge (Parkinson et al., 2002; Simpson, 1980; Webb, 1976; Wüster, Salomão, et al., 2002)
<i>B. lanceolatus</i>	AS	1.91 [0.2–6.43]	Water dispersal with island speciation (Wüster, Thorpe, et al., 2002)

Note: The *B. jararacussu* clade is object of study of an ongoing research, and it will not be presented herein.

Abbreviations: AS, Antillean subregion; BS, Brazilian subregion; CS, Chacoan subregion; SATZ, South American Transition.

5 | CONCLUSION

We had predicted a priori that (a) the diversification in *Bothrops* is ancient and (b) the geographic pattern of speciation was most likely unidirectional, having a north to south movement and diversification pattern that was not correlated with geoclimatic events. Our results showed a complex pattern of diversification for *Bothrops* in the Neotropics, with the uplift of the Andes likely playing an important role in the early diversification during the Pliocene-Miocene, and multifaceted past geoclimatic events helping to explain the diversification of younger clades of the genus along forested and open landscapes in the Neotropics during the Pleistocene-Pliocene. It is probable that some of the simpler biogeographic models proposed for the Neotropics are applicable at a more local scale but have limited applicability at such a vast regional scale. Our findings warrant further investigation of diversification and historical biogeographic studies across speciose, widely distributed clades.

TABLE 1 Estimated divergence times (in millions of years), ancestral area and putative events for evolution of *Bothrops* clades

ACKNOWLEDGEMENTS

We are grateful to the Instituto Vital Brazil for providing tissue samples, and Dr. Davor Vrcibradic (UNIRIO) and Dr. Hélio R. Silva (UFFRJ) for comments that greatly improved the manuscript. Two anonymous reviewers provided detailed and helpful comments on first draft of the manuscript. B.H. thanks CAPES and the Biodiversity and Evolutionary Biology Graduate Program of the Federal University of Rio de Janeiro for the financial support. T.B.G. thanks to FAPESP (2013/04170-8) and State University of Maranhão (UEMA) for the Senior Researcher fellowship.

CONFLICT OF INTEREST

We have no competing interests.

AUTHOR CONTRIBUTIONS

BH conceived the paper main goals; BH and TBG collected the data, BH and TBG performed the analyses; and BH led the writing with contribution from TBG, PC and JM in the interpretation and discussion of the results.

ORCID

Breno Hamdan  <https://orcid.org/0000-0001-7118-6162>
 Thaís B. Guedes  <https://orcid.org/0000-0003-3318-7193>
 Jane Melville  <https://orcid.org/0000-0002-9994-6423>

DATA AVAILABILITY STATEMENT

The data is available in Supporting Information (see below).

REFERENCES

- Albino, A. M., & Montalvo, C. I. (2006). Snakes from the Cerro Azul Formation (Upper Miocene), central Argentina, with a review of fossil viperids from South America. *Journal of Vertebrate Paleontology*, *26*, 581–587. [https://doi.org/10.1671/0272-4634\(2006\)26\[581:S-FTCAF\]2.0.CO;2](https://doi.org/10.1671/0272-4634(2006)26[581:S-FTCAF]2.0.CO;2)
- Alencar, L. R. V., Quental, T. B., Grazziotin, F. G., Alfaro, M. L., Martins, M., Venzon, M., & Zaher, H. (2016). Diversification in vipers: Phylogenetic relationships, time of divergence and shifts in speciation rates. *Molecular Phylogenetics and Evolution*, *105*, 50–62. <https://doi.org/10.1016/j.ympev.2016.07.029>
- Antonelli, A., Nylander, J. A. A., Persson, C., & Sanmartín, I. (2009). Tracing the impact of the Andean uplift on Neotropical plant evolution. *Proceedings of the National Academy of Sciences of the USA*, *106*, 9749–9754. <https://doi.org/10.1073/pnas.0811421106>
- Antonelli, A., & Sanmartín, I. (2011). Why are there so many plant species in the Neotropics? *Taxon*, *60*, 403–414. <https://doi.org/10.1002/tax.602010>
- Aragon, E., Goin, F. J., Aguilera, Y. E., Woodburne, M. O., Carlini, A. A., & Roggiero, M. F. (2011). Palaeogeography and palaeoenvironments of northern Patagonia from the Late Cretaceous to the Miocene: The Palaeogene Andean gap and the rise of the North Patagonian High Plateau. *Biological Journal of the Linnean Society*, *103*, 305–315. <https://doi.org/10.1111/j.1095-8312.2011.01684.x>
- Arévalo, E., Davis, S. K., & Sites, J. W. Jr (1994). Mitochondrial DNA sequence divergence and phylogenetic relationships among eight chromosome races of the *Sceloporus grammicus* complex (Phrynosomatidae) in central Mexico. *Systematic Biology*, *43*, 387–418. <https://doi.org/10.2307/2413675>
- Arteaga, A., Pyron, R. A., Peñafiel, N., Romero-Barreto, P., Culebras, J., Bustamante, L., ... Guayasamin, J. M. (2016). Comparative phylogeography reveals cryptic diversity and repeated patterns of cladogenesis for amphibians and reptiles in Northwestern Ecuador. *PLoS ONE*, *11*, e0151746. <https://doi.org/10.1371/journal.pone.0151746>
- Bacon, C. D., Molnar, P., Antonelli, A., Crawford, A. J., Montes, C., & Vallejo-Pareja, M. C. (2016). Quaternary glaciation and the Great American Biotic Interchange. *The Geological Society of America*, *44*, 375–378. <https://doi.org/10.1130/G37624.1>
- Barbo, F. E., Gasparini, J. L., Almeida, A. P., Zaher, H., Grazziotin, F. G., Gusmão, R. B., ... Sawaya, R. J. (2016). Another new and threatened species of lancehead genus *Bothrops* (Serpentes, Viperidae) from Ilha dos Franceses, Southeastern Brazil. *Zootaxa*, *4097*, 511–529. <https://doi.org/10.11646/zootaxa.4097.4.4>
- Barbo, F. E., Grazziotin, F. G., Sazima, I., Martins, M., & Sawaya, R. J. (2012). A new and threatened insular species of lancehead from Southeastern Brazil. *Herpetologica*, *68*, 418–429. <https://doi.org/10.1655/HERPETOLOGICA-D-12-00059.1>
- Brower, A. V. Z. (1994). Rapid morphological radiation and convergence among races of the butterfly *Heliconius erato* inferred from patterns of mitochondrial DNA evolution. *Proceedings of the National Academy of Sciences of the United States of America*, *91*, 6491–6495. <https://doi.org/10.1073/pnas.91.14.6491>
- Bryson, R. W., García-Vázquez, U. O., & Riddle, B. R. (2011). Phylogeography of Middle American gophersnakes: Mixed responses to biogeographical barriers across the Mexican Transition Zone. *Journal of Biogeography*, *38*, 1570–1584. <https://doi.org/10.1111/j.1365.2699.2011.02508.x>
- Buffetaut, E., Marandat, B., & Sigé, B. (1986). Découverte de dents de deinonychosaures (Saurischia, Theropoda) dans le Crétacé supérieur du Sud de la France [Discovery of deinonychosaur teeth (Saurischia, Theropoda) in the Upper Cretaceous of southern France]. *Comptes Rendus De L'académie Des Sciences À Paris, Série II*, *303*, 1393–1396.
- Campbell, J. A., & Lamar, W. W. (1989). *The venomous reptiles of Latin America*. London, UK: Comstock.
- Campbell, J. A., & Lamar, W. W. (2004). *Venomous reptiles of Western Hemisphere*. Ithaca, NY: Comstock Publishing Associates.
- Carrasco, P. A., Grazziotin, F. G., Farfán, R. S. C., Koch, C., Ochoa, J. A., Scrocchi, G. J., ... Chaparro, J. C. (2019). A new species of *Bothrops* (Serpentes: Viperidae: Crotalinae) from Pampas del Heath, southeastern Peru, with comments on the systematics of *Bothrops neuwiedi* group. *Zootaxa*, *4565*, 301–344. <http://dx.doi.org/10.11646/zootaxa.4565.3.1>
- Carrasco, P. A., Harvey, M. B., & Muñoz Saravia, A. (2009). The rare Andean pitviper *Rhinocerocephis jonathani* (Serpentes: Viperidae: Crotalinae): Redescription with comments on its systematics and biogeography. *Zootaxa*, *2283*, 1–15.
- Carrasco, P. A., Leynaud, G. C., & Scrocchi, G. J. (2010). Redescription of the southernmost snake species *Bothrops ammodytoides* (Serpentes: Viperidae: Crotalinae). *Amphibia-Reptilia*, *31*, 323–338. <https://doi.org/10.1163/156853810791769491>
- Carrasco, P. A., Mattoni, C. I., Leynaud, G. C., & Scrocchi, G. J. (2012). Morphology, phylogeny and taxonomy of South American bothropoid pitvipers (Serpentes: Viperidae). *Zoologica Scripta*, *41*, 109–124. <https://doi.org/10.1111/j.1463-6409.2011.00511.x>
- Carrasco, P. A., Venegas, P. J., Chaparro, J. C., & Scrocchi, G. J. (2016). Nomenclatural instability in the venomous snakes of the *Bothrops* complex: implications in toxinology and public health. *Toxicon*, *119*, 122–128. <https://doi.org/10.1016/j.toxic.2016.05.014>
- Cisneros-Heredia, D. F., Borja, M. O., Proaño, D., & Touzet, J. M. (2006). Distribution and natural history of the Ecuadorian toad-headed pitvipers of the genus *Bothrocophias* (Squamata: Serpentes: Viperidae: Crotalinae). *Herpetozoa*, *19*, 17–26.
- Colston, T. J., Grazziotin, F. G., Shepard, D. B., Vitt, L. J., Colli, G. R., & Henderson, R. W. (2013). Molecular systematics and historical biogeography of tree boas (*Corallus* spp.). *Molecular Phylogenetics and Evolution*, *66*, 953–959. <https://doi.org/10.1016/j.ympev.2012.11.027>
- Conn, J. E., & Mirabello, L. (2007). The biogeography and population genetics of neotropical vector species. *Heredity*, *99*, 245–256. <https://doi.org/10.1038/sj.hdy.6801002>
- Darriba, D., Taboada, G. L., Doallo, R., & Posada, D. (2012). jModelTest 2: More models, new heuristics and parallel computing. *Nature Methods*, *9*, 772. <https://doi.org/10.1038/nmeth.2109>
- Dick, C. W., Abdul-Salim, K., & Bermingham, E. (2003). Molecular systematic analysis reveals cryptic tertiary diversification of a widespread tropical rain forest tree. *American Naturalist*, *162*, 691–703. <https://doi.org/10.1086/379795>

- Elias, M., Joron, M., Willmott, K., Silva-Brandão, K. L., Kaiser, V., Arias, C. F., ... Jiggins, C. D. (2009). Out of the Andes: Patterns of diversification in clearwing butterflies. *Molecular Ecology*, *18*, 1716–1729. <https://doi.org/10.1111/j.1365-294X.2009.04149.x>
- Felsenstein, J. (1985). Confidence limits on phylogenies: An approach using the bootstrap. *Evolution*, *39*, 783–791. <https://doi.org/10.1111/j.1558-1985.tb00420.x>
- Fenwick, A. M., Greene, H. W., & Parkinson, C. L. (2012). The serpent and the egg: unidirectional evolution of reproductive modes in vipers? *Journal of Zoological Systematics and Evolutionary Research*, *50*, 59–66. <https://doi.org/10.1111/j.1439-0469.2011.00646.x>
- Fenker, J., Tedeschi, L. G., Pyron, R. A., & Nogueira, C. C. (2014). Phylogenetic diversity, habitat loss and conservation in South American pitvipers (Crotalinae: *Bothrops* and *Bothrocophias*). *Diversity and Distributions*, *20*, 1108–1119. <https://doi.org/10.1111/ddi.12217>
- Fenwick, A. M., Gutberlet, R. L. Jr, Evans, J. A., & Parkinson, C. L. (2009). Morphological and molecular evidence for phylogeny and classification of South American pitvipers, genera *Bothrops*, *Bothriopsis*, and *Bothrocophias* (Serpentes: Viperidae). *Zoological Journal of the Linnean Society*, *156*, 617–640. <https://doi.org/10.1111/j.1096-3642.2008.00495.x>
- Folguera, A., Orts, D., Spagnuolo, M., Vera, E. R., Sagripanti, L., Ramos, M. E., & Ramos, V. A. (2011). A review of Late Cretaceous to Quaternary palaeogeography of the southern Andes. *Biological Journal of the Linnean Society*, *103*, 250–268. <https://doi.org/10.1111/j.1095-8312.2011.01687.x>
- George, W., & Vincent, S. (1977). Report of field meeting Walton-on-the-Naze and Wrabness, Essex, 2.X, 1976 with notes on the London Clay of Walton. *Tertiary Research*, *1*, 83–90.
- Grau, E. T., Pereira, S. L., Silveira, L. F., Wajntal, A., & Höfling, E. (2005). Molecular phylogenetics and biogeography of Neotropical piping guans (Aves: Galliformes): *Pipile* Bonaparte, 1856 is synonym of *Aburria* Reichenbach, 1853. *Molecular Phylogenetics and Evolution*, *35*, 637–645. <https://doi.org/10.1016/j.ympev.2004.12.004>
- Grazziotin, F. G., Monzel, M., Echeverrigaray, S., & Bonatto, S. L. (2006). Phylogeography of the *Bothrops jararaca* complex (Serpentes: Viperidae): Past fragmentation and island colonization in the Brazilian Atlantic Forest. *Molecular Ecology*, *15*, 3969–3982. <https://doi.org/10.1111/j.1365-294X.2006.03057.x>
- Gregory-Wodzicki, K. M. (2000). Uplift history of the Central and Northern Andes: A review. *Geological Society of America Bulletin*, *112*, 1091–1105. [https://doi.org/10.1130/0016-7606\(2000\)112<1091:UHOTCA>2.CO;2](https://doi.org/10.1130/0016-7606(2000)112<1091:UHOTCA>2.CO;2)
- Guarnizo, C. E., Werneck, F. P., Giugliano, L. G., Santos, M. G., Fenker, J., Sousa, L., & Colli, G. R. (2016). Cryptic lineages and diversification of an endemic anole lizard (Squamata, Dactyloidae) of the Cerrado hotspot. *Molecular Phylogenetics and Evolution*, *94*, 279–289. <https://doi.org/10.1016/j.ympev.2015.09.005>
- Guedes, T. B., Nogueira, C., & Marques, O. A. V. (2014). Diversity, natural history, and geographic distribution of snakes in the Caatinga, Northeastern Brazil. *Zootaxa*, *3863*, 1–93. <https://doi.org/10.11646/zootaxa.3863.1.1>
- Guedes, T. B., Sawaya, R. J., Zizka, A., Laffan, S., Faurby, S., Pyron, R. A., ... Antonelli, A. (2018). Patterns, biases, and prospects in the distribution and diversity of Neotropical snakes. *Global Ecology and Biogeography*, *27*, 14–21. <https://doi.org/10.1111/geb.12679>
- Guindon, S., & Gascuel, O. (2003). A simple, fast, and accurate algorithm to estimate large phylogenies by maximum likelihood. *Systematic Biology*, *52*, 696–704. <https://doi.org/10.1080/10635150390235520>
- Haffer, J. (1969). Speciation in Amazonian forests birds. *Science*, *165*(3889), 131–137. <https://doi.org/10.1126/science.165.3889.131>
- Haffer, J. (2008). Hypotheses to explain the origin of species in Amazonia. *Brazilian Journal of Biology*, *68*, 917–947. <https://doi.org/10.1126/science.215.4538.1351>
- Hamdan, B., Pereira, A. G., Loss-Oliveira, L., Rödder, D., & Schrago, C. G. (2017). Evolutionary analysis of *Chironius* snakes unveils cryptic diversity and provides clues to diversification in the Neotropics. *Molecular Phylogenetics and Evolution*, *116*, 108–119. <https://doi.org/10.1016/j.ympev.2017.08.004>
- Harvey, M. B. (1994). A new montane pitviper (Viperidae: *Bothrops*) from Cochabamba, Bolivia. *Proceedings of the Biological Society of Washington*, *107*, 60–66.
- Head, J. J. (2015). Fossil calibration dates for molecular phylogenetic analysis of snakes 1: Serpentes, Alethinophidia, Boidae, Pythonidae. *Palaeontologia Electronica*, *18.1.6FC*, 1–17. <https://doi.org/10.26879/487>
- Head, J. J., Mahlow, K., & Müller, J. (2016). Fossil calibration dates for molecular phylogenetic analysis of snakes 2: Caenophidia, Colubroidea, Elapoidea, Colubridae. *Palaeontologia Electronica*, *19.2.2FC*, 1–21. <https://doi.org/10.26879/625>
- Hedges, S. B. (1996). The origin of West Indian amphibians and reptiles. In R. Powell & R. W. Henderson (Eds.), *Contributions to West Indian herpetology: A tribute to Albert Schwartz* (pp. 95–128). Ithaca, NY: Society for the Study of Amphibians and Reptiles.
- Hooghiemstra, H., & Cleef, A. M. (1995). Pleistocene climatic change and environmental and generic dynamics in the north Andean montane forest and paramo. In S. P. Churchill, H. Balslev, E. Forero, & J. L. Luyeyn (Eds.), *Biodiversity and conservation of Neotropical Montane Forests* (pp. 35–49). New York, NY: Botanical Garden.
- Hoorn, C., Wesselingh, F. P., ter Steege, H., Bermudez, M. A., Mora, A., Sevink, J., ... Antonelli, A. (2010). Amazonia through time: Andean uplift, climate change, landscape evolution and biodiversity. *Science*, *330*, 927–931. <https://doi.org/10.1126/science.1194585>
- Inoue, J., Donoghue, P. C., & Yang, Z. (2010). The impact of the representation of fossil calibrations on Bayesian estimation of species divergence times. *Systematic Biology*, *59*, 74–89. <https://doi.org/10.1093/sysbio/syp078>
- Irwin, D. M., Kocher, T. D., & Wilson, A. C. (1991). Evolution of the cytochrome b gene of mammals. *Journal of Molecular Evolution*, *32*, 128–144. <https://doi.org/10.1007/BF02515385>
- Jacobs, B. F. (2004). Palaeobotanical studies from tropical Africa: Relevance to evolution of forest, woodland and savannah biomes. *Philosophical Transactions of the Royal Society B*, *359*, 1573–1583. <https://doi.org/10.1098/rstb.2004.1533>
- Katoh, K., & Standley, D. M. (2013). MAFFT Multiple Sequence Alignment Software Version 7: Improvements in Performance and Usability. *Molecular Biology and Evolution*, *30*, 772–780. <https://doi.org/10.1093/molbev/mst010>
- Knight, A., & Mindell, D. P. (1994). The phylogenetic relationship of Colubrinae, Elapidae and Viperidae and the evolution of front-fanged venom systems in snakes. *Copeia*, *1994*, 1–9. <https://doi.org/10.2307/1446664>
- Kocher, T. D., Thomas, W. K., Meyer, A., Edwards, S. V., Pääbo, S. F., Villablanca, R. X., & Wilson, A. C. (1989). Dynamics of mtDNA evolution in animals: Amplification and sequencing with conserved primers. *Proceedings of the National Academy of Sciences of the*

- United States of America*, 86, 6196–6200. <https://doi.org/10.1073/pnas.86.16.6196>
- Lira-da-Silva, R. M., Mise, Y. F., Casais-e-Silva, L. L., Ulloa, J., Hamdan, B., & Brazil, T. K. (2009). Serpentes de Importância Médica do Nordeste do Brasil. *Gazeta Médica Da Bahia*, 79, 7–20.
- Lohmann, L. G., Bell, C. D., Calió, M. F., & Winkworth, R. C. (2013). Pattern and timing of biogeographical history in the Neotropical tribe Bignoniaceae (Bignoniaceae). *Botanical Journal of the Linnean Society*, 171, 154–170. <https://doi.org/10.1111/j.1095-8339.2012.01311.x>
- Machado, T., Silva, V. X., & Silva, M. J. J. (2014). Phylogenetic relationships within *Bothrops neuwiedi* clade (Serpentes, Squamata): Geographically highly-structured lineages, evidence of introgressive hybridization and Neogene/Quaternary diversification. *Molecular Phylogenetics and Evolution*, 71, 1–14. <https://doi.org/10.1016/j.ympev.2013.10.003>
- Marshall, L. G., Webb, S. D., Sepkoski, J. J. Jr, & Raup, D. M. (1982). Mammalian evolution and the great American interchange. *Science*, 215, 1351–1357. <https://doi.org/10.1126/science.215.4538.1351>
- Martins, M., Araujo, M. S., Sawaya, R. J., & Nunes, R. (2001). Diversity and evolution of macrohabitat use, body size and morphology in a monophyletic group of Neotropical pitvipers (*Bothrops*). *Journal of Zoology*, 254, 529–553. <https://doi.org/10.1017/S0952836901001030>
- Miller, M. A., Pfeiffer, W., & Schwartz, T. (2010). Creating the CIPRES Science Gateway. *Gateway Computing Environments Workshop*, 11705685, 1–8. <https://doi.org/10.1109/GCE.2010.5676129>
- Morrone, J. J. (2014). Biogeographical regionalisation of the Neotropical region. *Zootaxa*, 3782, 1–110. <https://doi.org/10.11646/zootaxa.3782.1.1>
- Myers, N., Mittermeier, R. A., Mittermeier, C. G., Fonseca, G. A. B., & Kent, J. (2000). Biodiversity hotspots for conservation priorities. *Nature*, 403, 853–858. <https://doi.org/10.1038/35002501>
- Ospina-L., A. M. (2017). *Bothrops punctatus* (García 1896). *Catálogo De Anfíbios Y Reptiles De Colombia*, 3, 25–30.
- Palumbi, S. R., Martin, A., Romano, S., McMillan, W. O., Stice, L., & Grabowski, G. (1991). *The simple fool's guide to PCR, version 2.0*. Privately published document compiled by S. Palumbi, Department of Zoology and Kewalo Marine Laboratory. Honolulu, HI: University of Hawaii.
- Parkinson, C. L., Campbell, J. A., & Chippindale, P. T. (2002). Multigene phylogenetic analyses of pitvipers; with comments on the biogeographical history of the group. In G. W. Schuett, M. Höggren, M. E. Douglas, & H. W. Greene (Eds.), *Biology of the vipers* (pp. 93–110). Eagle Mountain, Utah: Eagle Mountain Publishing.
- Parmley, D., & Holman, J. A. (2007). Earliest fossil record of a Pigmy Rattlesnake (Viperidae: *Sistrurus Garmani*). *Journal of Herpetology*, 41, 141–144. [https://doi.org/10.1670/0022-1511\(2007\)41\[141:E-FROAP\]2.0.CO;2](https://doi.org/10.1670/0022-1511(2007)41[141:E-FROAP]2.0.CO;2)
- Parmley, D., & Hunter, K. B. (2010). Fossil snakes of the Clarendonian (late Miocene) Pratt Slide local fauna of Nebraska, with the description of a new natricine colubrid. *Journal of Herpetology*, 44, 526–543. <https://doi.org/10.1670/09-248.1>
- Plana, V. (2004). Mechanisms and tempo of evolution in the African Guineo-Congolian rainforest. *Philosophical Transactions of the Royal Society B*, 359, 1585–1594. <https://doi.org/10.1098/rstb.2004.1535>
- Ramírez, S. R., Roubik, D. W., Skov, C., & Pierce, N. E. (2010). Phylogeny, diversification patterns and historical biogeography of euglossine orchid bees (Hymenoptera: Apidae). *Biological Journal of the Linnean Society*, 100, 552–572. <https://doi.org/10.1111/j.1095-8312.2010.01440.x>
- Raymo, M. E., Hearty, P., De Conto, R., O'Leary, M., Dowsett, H. J., Robinson, M. M., & Mitrovica, J. X. (2009). PLIOMAX: Pliocene maximum sea level project. *Pages News*, 17, 58–59. <https://doi.org/10.22498/pages.17.2.58>
- Ree, R. H., Moore, B. R., Webb, C. O., & Donoghue, M. J. (2005). A likelihood framework for inferring the evolution of geographic range on phylogenetic trees. *Evolution*, 59, 2299–2311. <https://doi.org/10.1111/j.0014-3820.2005.tb00940.x>
- Ribas, C. C., Aleixo, A., Nogueira, A. C. R., Miyaki, C. Y., & Cracraft, J. (2012). A palaeobiogeographic model for biotic diversification within Amazonia over the past three million years. *Proceeding of the Royal Society B*, 279, 681–689. <https://doi.org/10.1098/rspb.2011.1120>
- Rull, V. (2011). Neotropical biodiversity: Timing and potential drivers. *Trends in Ecology and Evolution*, 26, 508–513. <https://doi.org/10.1016/j.tree.2011.05.011>
- Saldarriaga-Córdoba, M., Parkinson, C. L., Daza, J. M., Wüster, W., & Sasa, M. (2017). Phylogeography of the Central American lancehead *Bothrops asper* (Serpentes: Viperidae). *PLoS ONE*, 12, e0187969. <https://doi.org/10.1371/journal.pone.0187969>
- Sauquet, H., Ho, S. Y. W., Gandolfo, M. A., Jordan, G. J., Wilf, P., Cantrill, D. J., ... Udovicic, F. (2012). Testing the Impact of Calibration on Molecular Divergence Times Using a Fossil-Rich Group: The Case of *Nothofagus* (Fagales). *Systematic Biology*, 61, 289–313. <https://doi.org/10.1093/sysbio/syr116>
- Savage, J. M. (1966). The origins and history of the Central American Herpetofauna. *Copeia*, 1966, 719–766. <https://doi.org/10.2307/1441404>
- Scanferla, C. A., & Nenda, S. J. (2005). El registro más antiguo del género *Bothrops* (Serpentes, Viperidae), proveniente del Pleistoceno inferior a medio de Argentina. *Revista Del Museo Argentino Di Ciencias Naturales*, 7, 177–181.
- Schluter, D. (2000). *The ecology of adaptive radiation*. Oxford, UK: Oxford University Press.
- Silva, V. X., & Rodrigues, M. T. (2008). Taxonomic revision of the *Bothrops neuwiedi* complex (Serpentes, Viperidae) with description of a new species. *Phyllomedusa*, 7, 45–90. <https://doi.org/10.11606/issn.2316-9079.v7i1p45-90>
- Simpson, G. G. (1980). *Splendid isolation: The curious history of South American mammals*. New Haven, Connecticut: Yale University Press.
- Smith, B. T., McCormack, J. E., Cuervo, A. M., Hickerson, M. J., Aleixo, A., Cadena, C. D., ... Brumfield, R. T. (2014). The drivers of tropical speciation. *Nature*, 515, 406–409. <https://doi.org/10.1038/nature13687>
- Tamura, K., Battistuzzi, F. U., Billing-Ross, P., Murillo, O., Filipski, A., & Kumar, S. (2012). Estimating divergence times in large molecular phylogenies. *Proceedings of the National Academy of Sciences of the United States of America*, 109, 19333–19338. <https://doi.org/10.1073/pnas.1213199109>
- Timms, J., Chaparro, J. C., Venegas, P. J., Salazar-Valenzuela, D., Scrocchi, G., Cuevas, J., ... Carrasco, P. A. (2019). A new species of pitviper of the genus *Bothrops* (Serpentes: Viperidae: Crotalinae) from the Central Andes of South America. *Zootaxa*, 4656(1), 099–120. <https://doi.org/10.11646/zootaxa.4656.1.4>
- Turci, L. C. B., Albuquerque, S., Bernarde, P. S., & Miranda, D. B. (2009). Uso do hábitat, atividade e comportamento de *Bothriopsis bilineatus* e de *Bothrops atrox* (Serpentes: Viperidae) na floresta do

- Rio Moa, Acre, Brasil. *Biota Neotropica*, 9, 197–206. <https://doi.org/10.1590/S1676-06032009000300020>
- Uetz, P., Freed, P., & Hošek, J. (2018). *The reptile database*. Retrieved from <http://www.reptile-database.org>
- Vidal, N., Lecomte, G. C., Vié, J. C., & Gasc, J. P. (1997). Molecular systematic of pitvipers: Paraphyly of the *Bothrops* complex. *Evolution*, 320, 95–101. [https://doi.org/10.1016/S0764-4469\(99\)80091-2](https://doi.org/10.1016/S0764-4469(99)80091-2)
- Wallace, A. R. (1852). On the monkeys of the Amazon. *Proceeding of the Zoological Society of London*, 20, 107–110.
- Wallach, V., Williams, K. L., & Boundy, J. (2014). *Snakes of the world: A catalogue of living and extinct species*. Boca Raton, FL: Taylor and Francis: CRC Press.
- Webb, S. D. (1976). Mammalian faunal dynamics of the Great American Biotic Interchange. *Paleobiology*, 2, 216–234.
- Weigt, L. A., Crawford, A. J., Rand, A. S., & Ryan, M. J. (2005). Biogeography of the tungara frog, *Physalaemus pustulosus*: A molecular perspective. *Molecular Ecology*, 14(12), 3857–3876.
- Werman, S. D. (2005). Hypotheses on the historical biogeography of bothropoid pitvipers and related genera of the Neotropics. In M. A. Donnelly, B. I. Crother, C. Guyer, M. H. Wake, & M. E. White (Eds.), *Ecology and evolution in the tropics: A herpetological perspective* (pp. 306–365). Chicago, IL: University of Chicago Press.
- Werneck, F. P. (2011). The diversification of eastern South American open vegetation biomes: Historical biogeography and perspectives. *Quaternary Science Review*, 30, 1630–1648. <https://doi.org/10.1016/j.quascirev.2011.03.009>
- Werneck, F. P., Leite, R. N., Geurgas, S. R., & Rodrigues, M. T. (2015). Biogeographic history and cryptic diversity of saxicolous Tropicuridae lizards endemic to the semiarid Caatinga. *BMC Evolutionary Biology*, 15, 1–24. <https://doi.org/10.1186/s12862-015-0368-3>
- White, T. E. (1942). The Lower Miocene mammal fauna of Florida. *Bulletin of the Museum of Comparative Zoology*, 92, 1–49.
- Wüster, W., Peppin, L., Pook, C. E., & Walker, D. E. (2008). A nesting of vipers: Phylogeny and historical biogeography of the Viperidae (Squamata: Serpentes). *Molecular Phylogenetics and Evolution*, 49(2008), 445–459. <https://doi.org/10.1016/j.ympev.2008.08.019>
- Wüster, W., Salomão, M. G., Duckett, G. J., Thorpe, R. S., & BBBSP (1999). Mitochondrial DNA phylogeny of the *Bothrops atrox* species complex (Squamata: Serpentes: Viperidae). *Kaupia*, 8, 135–144.
- Wüster, W., Salomão, M. G., Quijada-Mascareñas, J. A., Thorpe, R. S., & BBBSP (2002). Origins and evolution of the South American pitvipers fauna: Evidence from mitochondrial DNA sequence analysis. In G. W. Schuett, M. Höggren, M. E. Douglas, & H. W. Greene (Eds.), *Biology of the vipers* (pp. 111–129). Eagle Mountain UT: Eagle Mountain Publishing.
- Wüster, W., Thorpe, R. S., Salomão, M. D. G., Thomas, L., Puerto, G., Theakston, R. D. G., & Warrell, D. A. (2002). Origin and phylogenetic position of the Lesser Antillean species of *Bothrops* (Serpentes Viperidae): Biogeographic and medical implications. *Bulletin of the Natural History Museum of London*, 68, 101–106. <https://doi.org/10.1017/S0968047002000110>
- Yu, Y., Harris, A. J., Blair, C., & He, X. J. (2015). RASP (Reconstruct Ancestral State in Phylogenies): A tool for historical biogeography. *Molecular Phylogenetics and Evolution*, 87, 46–49. <https://doi.org/10.1016/j.ympev.2015.03.008>
- Zamudio, K. R., & Greene, H. W. (1997). Phylogeography of the bushmaster (*Lachesis muta*: Viperidae): Implications for neotropical biogeography, systematics, and conservation. *Biological Journal of the Linnean Society*, 62, 421–442. <https://doi.org/10.1111/j.1095-8312.1997.tb01634.x>

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

How to cite this article: Hamdan B, Guedes TB, Carrasco PA, Melville J. A complex biogeographic history of diversification in Neotropical lancehead pitvipers (Serpentes, Viperidae). *Zool Scr.* 2019;00: 1–14. <https://doi.org/10.1111/zsc.12398>